

Building Vocal Fold Solid Models from CAD models created from CT and Histological data

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Abstract

Quantification of vocal fold geometry is necessary for the development of anatomically realistic and consistently defined experimental/computational models of the glottic and subglottic regions. Such models will facilitate the study of the influence of the subglottis in voice production. This Technical memo is provided primarily as a tutorial of one method to create physical models from CT data. Download updates to this memo can be downloaded at <http://www.nrlld.org/>.

Keywords: Laryngeal Cartilage, CAD model, MRI, vocal folds

1. Introduction

Understanding the morphology of the laryngeal framework is a crucial step in mapping laryngeal function. Further, accurate dimensions of all structures of the larynx is necessary to produce reliable quantitative biomechanical modeling of voice disorders. Particularly important are dimensions of those structures which describe function in mechanical terms (e.g., laryngeal muscle orientation, cartilage length/width, and joint size/range).

Magnetic Resonance Imaging (MRI) and X-ray computed tomography (CT) scans have become useful tools in obtaining anatomical information in both live subjects and donated samples. In the final form, such images can be used to create geometric shapes. However, there are difficulties with each of these methods. CT only clearly captures boundary distinctions (soft tissue to hard tissue, or tissue to air). On the other hand, MRI can give more tissue variation differences but not at high resolutions. Finally, while histological images achieve better distinction between differing tissue types, the freezing and slicing procedures may introduce some geometric distortion.

Because of the increased use of these techniques, getting from the imaging to either virtual CAD or physical models is important. The purpose of this report is to discuss a method of creating solid models from MRI, CT and histological images.

2. Histological Image Analysis towards a Finite Element Model

In this section we demonstrate the feasibility of generating a finite element mesh using a vocal fold histological section. Consider the histological section of the vocal fold shown in Fig. 1a where the three layers are distinguishable. The boundary of the three layers was traced with image editing software, using knowledge of the

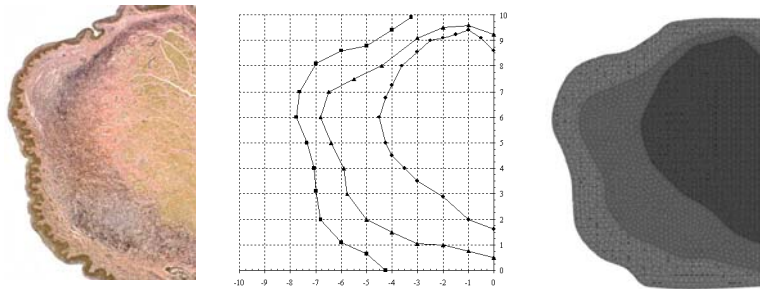


Figure 1: Process of converting histological vocal fold section to finite element model.

anatomy and by adjusting image contrast to help determine the boundary location (Fig. 1b). The boundaries approximately corresponded to: (1) the epithelial lining along the medial surface; (2) the interface between the superficial and intermediate lamina propria layers; and (3) the thyroarytenoid muscle boundary. This resulted in three layers, corresponding to the cover, ligament, and body (muscle). To generate a finite element mesh, the point coordinates were imported into the commercial code ADINA, and splines were used to connect the various points. The three layers were meshed individually, resulting in a three-layer finite element model, shown in Fig. 1c. Extraction of the image data from two to three dimensions using multiple images is a straightforward extension of the MRI-based methods discussed below.

3. Creating Virtual Models

Consider a series of histological slides with extraction of features as discussed in the previous section. To create the virtual models, series of point data for each tissue layer (for example, a series of curves defining the thyroarytenoid muscle) were imported into Pro/Engineer and blended together. Figure 2 illustrates the creation of such a three-dimensional virtual model. For this particular model, a series of curves

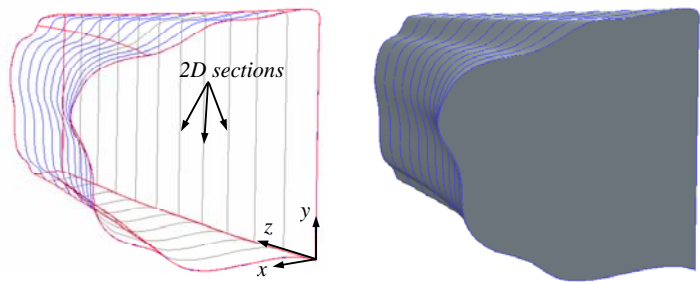


Figure 2: Example of creating three-dimensional virtual model from two-dimensional sections. Left: Series of sections, with each section lying in the x-y plane. Right: Virtual model generated by blending sections

was first created. Each of these curves represented a two-dimensional section in the x-y plane; the curves were slightly offset in the z-direction. Note that the first of these curves (at the $z = 0$ plane) was taken from the histological image analysis shown in Fig. 1a; the curves offset in the z-direction were modifications of the first curve, generated in this case purely for illustration purposes. The curves were imported into Pro/Engineer and a connected virtual three-dimensional model was generated by blending the series of two-dimensional sections. The solid model on the right side of Fig. 2 were then used in Modeling or Rapid Prototyping applications.

Additional detail can be obtained and included. Consider the series of curves generated using the digitized histological images from Figure 1 delineating the various tissue layers in the vocal fold. Similar curves can be obtained from each histological (or other) image and imported into Pro/Engineer and converted to three-dimensional virtual models, similar to the manner used in Fig. 2. This will yield virtual models of each layer of tissue for each larynx sample. These models, as a whole or part, can be saved in a solid model format (e.g., IGES). The virtual tissue layer models from each larynx sample can then be assembled (also in Pro/Engineer) to create a complete virtual laryngeal model. This is illustrated in Fig. 3, where an assembly of three-dimensional virtual models of the vocal fold body, ligament, and cover layers is represented.

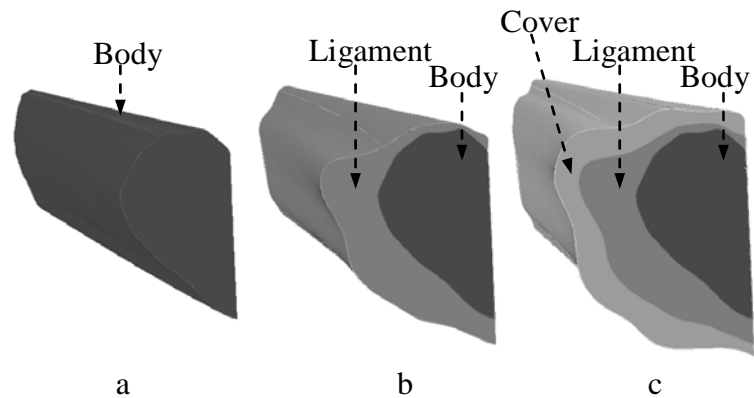


Figure 3: Three-dimensional virtual vocal fold model created from two-dimensional slices. (a) Body layer; (b) Body and ligament layers; (c) Body, ligament, and cover layers.

4. MRI to Solid Modeling

In this section, an actual example is given where a series of 128 MRI scans from one human larynx (see NRLD#000001) were used to provide the basic geometry. Using commercial MRI and three-dimensional software packages (Velocity2 and Pro/Engineer, respectively), the images were imported. Grayscale thresholding techniques were used to identify and extract three-dimensional representations of the vocal folds and of the cricoid, thyroid, and arytenoid cartilages. Scaling of the images was done in consultation with cartilage data from Hunter and Titze (2005). These images and eventual solid models were output as IGES files.

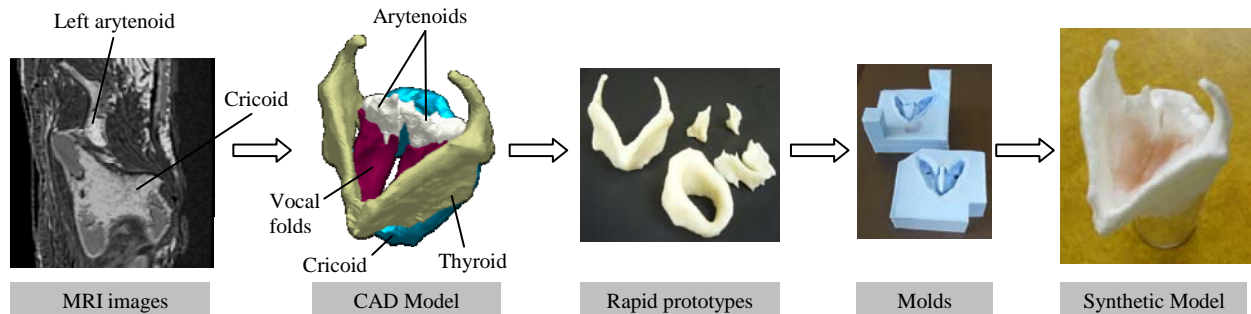


Figure 4: MRI-based synthetic larynx model fabrication process

Physical models of these features were fabricated using a computer-controlled rapid prototyping machine. From these models, molds were constructed. Cartilage models were made by pouring liquid plastic into the cartilage molds and allowing the plastic to cure. Liquid silicone was poured into the vocal folds mold and allowed to cure to create the vocal folds model. The vocal folds had a Young's modulus of approximately 2 kPa, similar to that of superficial vocal

fold tissue layers. The cartilages and vocal folds were fastened together using flexible silicone. The resulting synthetic “joints” were of sufficient stiffness so as to preserve cartilage framework positioning but to allow deformation when acted upon by an external force. A silicone membrane of desired geometry was also attached to simulate the conus elasticus.

Literature

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Use Agreement

The scripts, images and text are open to use by the public as a service and part of the National Resource of Laryngeal Data (supported by the National Institute of Deafness and other Communicative Disorders, and hosted by the National Center for Voice and Speech). The scripts, images, model and text enclosed in this memo and accompanying this memo are open to use by the public as a service of the NRLD. However, we ask the reader to respect the time and effort put into this manuscript and research. If the text, images, or included scripts are used, the user agrees to reference to this document, the NRLD, and the source of the original data. We also ask the users to consider contacting the original contributors of the data and give them the right of refusal to (1) participate on papers using the data and (2) have their supporting project acknowledged. The user agrees to freely share with the NLDR any extension software build on the data contained.

Revisions

1.0 Eric Hunter: Main document (March 2011)-- original raw data files and ability to read them.